**Western Blot SDS PAGE Protocol**

**Making Gel**

1. Set up Gel Caster based on PhosTsg Gel Making Protocol
2. Make 20% Glycerol, 0.5% 222-TriChloroethanol, 10% Acrylamide running gel based on gel formula
3. Make 10% Glycerol stacking gel, 4.5% acrylamide stacking gel based on gel formula

**Loading Gel & Running Electrophoresis**

1. Rinse wells of the stacking gel with DiWater then flick the caster with your wrists to get all the water out. The lanes of the gel should be sharp with no water. Do this 3 times to ensure the lanes are properly cleared
2. After the wells of the stacking gel have been rinsed and samples desired to analyse have been set on ice to thaw.

- Put gel-plates in buffer chamber with glass side facing out

- Clip the gel-plates to buffer chamber with the red clamps [longer side facing you].

- Mark the top of each well with a marker on each gel-plate

- Fill the buffer chamber between the gel-plates [bottom to top] with [Running Buffer](#Running_Buffer) [put in fridge to cool].

3. Vortex samples before loading

4. Fill the sample wells.

- Load 5 ul Protein Ladder into lane

- Load samples at desired amounts in desired order (1uL sample is usually used)

5. Take buffer chamber to fridge then fill the rest of the chamber with the remaining Running Buffer

6. Connect the buffer chamber with the voltage system (red on lid goes to red on chamber)

- **Run Gel for 20 minutes @ 20mA constant amp**

**- Then run gels for 2 hours @ 160 mV constant volt or until color portion is about an inch from the bottom of the gel**

7. If you are running more than two gels, start running the first two gels before you load the second two

**Transfer (after electrophoresis)**

1. Before electrophoresis ends:

* Make [transfer buffer](#Transfer_Buffer)
* If using **nitrocellulose membrane**: cut membrane the size of the filter paper (one for each gel). With the shiny side (inside of the roll) facing up, cut the bottom right corner of each membrane
* If using **PVDF membrane**: cut membrane the size of the filter paper (one for each gel). Let PVDF membrane rock in methanol for 20 minutes
* Put four pieces of filter paper inside the transfer cassettes then in the transfer chamber. Fill the transfer chamber halfway with transfer buffer

1. Take running chamber out of fridge and dump out running buffer
2. Take off red clips and slip gel-plates out of the chamber
3. Separate the glass plate from the aluminium plate (keeping the gel on the aluminium plate)
4. Cut the well off at the line between the stacking and running gel using a razor blade
5. Cut the gel just below the color line and then cut the bottom left corner off the get (I do this so I have a reference to which is side is the front of the gel in case it flips during washes).
6. If **stain free chemical** (222-Trichloroethanol) was used when making the gel then take gels directly off of aluminium plate and place on ChemiDoc with a few drops off water on the tray. Activate and image the gels with the “Stain Free Gel” application. This should be set to 45 seconds activation and auto optimal exposure.
7. Wash the gels and membranes (either nitrocellulose or PVDF) in transfer buffer for 5 minutes
8. Make a sandwich of the gel for transfer in this order;
   1. Black side of casket
   2. Sponge sheet
   3. Filter paper (thick blot paper)
   4. Gel
   5. Nitrocellulose membrane (shiny side of membrane facing down on the gel so the cut corners of the gel and the membrane line up) OR PVDF membrane
   6. Filter paper (thick blot paper)
   7. Sponge sheet
   8. Grey side of casket
   9. Place casket (clips side down) with the Black side of casket facing the back (where the negative (black) probe will be)
9. Put stir bar at the bottom of the transfer chamber (carefully)
10. Take running chamber to fridge, put on stir plate, and fill to max line with transfer buffer
11. Put lid on transfer chamber with black probe at the back of the chamber and red probe at the front of the chamber
12. Turn on stir apparatus so stir bar is moving transfer buffer around throughout the transfer process
13. Transfer samples onto membrane using the wet transfer chamber

* **For 1 gel: 1.5 hours @ 25mA constant amp**
* **For 2 gels: 1.5 hours @ 50mA constant amp**
* **For 4 gels: 1.5 hours @100mA constant amp**

**Blocking**

1. Before transfer has finished, make TBS-Tween + Milk
2. Save 10 mL of TBS-Tween + Milk for each gel you ran for primary antibodies
3. Save 10 mL more of TBS-Tween + Milk for each gel you ran for secondary antibodies
4. Remove transfer chamber from fridge and take out cassette
5. Carefully remove the membrane from the gel and place membranes in light sensitive black boxes with TBS-Tween and gently agitate for a few seconds
6. Pour out TBS-Tween that is on the membranes and pour in TBS-Tween + Milk to boxes with membranes. Let membranes rock for 1.5 hours
7. To confirm transfer:
   1. Carefully wash the transferred gels with water and place in a container with enough oriole stain to cover the gels. Let gels rock under tin foil for 1.5 hours then pour used oriole stain into used container and put gels in water to image. To image gels, take them to chemiDoc and use “oriole” stain application

**OR**

* 1. If you used the stain free chemical when making the gel, take the gel out of transfer cassette and image gels on chemidoc with a few drops of water on the tray using the “Stain Free Gel” application. No activation is required, so you can set the activation to zero with auto-optimal exposure

1. You may dispose of the transferred gels

**Primary Antibody**

1. After blocking, wash membraned in TBST 3 times for 5 minutes each
2. During the last wash, make primary antibody dilution using 10mL of TBST + Milk for each membrane (inquire about appropriate antibody dilution from research or previous experiments in this lab)
3. After washing membranes, put primary antibody in box with membranes and rock overnight in fridge.

**Secondary Antibody**

1. Take membranes out of fridge from overnight rocking and pour out primary antibody
2. Wash membranes in TBST 3 times for 5 minutes each
3. **Turn off lights to make secondary antibody dilution** using 10mL of TBST + Milk for each membrane (inquire about appropriate dilution from research or previous experiments in this lab)
4. Pour out last wash of TBST and pour in secondary antibody dilution to black light sensitive boxes
5. Places boxes with membranes on rocker under tin foil and let rock for 1 hour

**Imaging – All imaging must be done with lights off**

1. Wash membranes after secondary antibodies in TBST 3 times for 5 minutes each
2. If a HRP-Conjugated secondary antibody was used, use ECL signalling reagent to develop by combining 3mL of reagent A and 3mL of reagent B and placing membrane in mixture for 60 seconds before imaging **(ensure lights are turned off)** 
   * Take membrane out of reagent and place face down on chemidoc in room 531
   * Use chemiluminescent blot application to image membrane
3. If a Alexa Fluor conjugated secondary antibody was used, take membranes out of TBST and place between two pieces of filter paper and in a dark box to let dry for 10 minutes
   * Take membrane out of filter paper and place face down on chemidoc in room 518
   * Image with wavelength indicated on secondary antibody
4. You may want to get another total protein on the membrane if you used the stain free chemical by using the “stain free blot” application

\*\*After imaging you may want to perform a Ponceau total protein stain on the membranes if a total protein was not taken before transfer with the stain free gel. See Ponceau Stain Protocol for more information\*\*